EDITORIAL REVIEW

Heat shock proteins and innate immunity

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Heat shock proteins (hsp) have attracted considerable attention from immunologists over the last 20 years, and their interest has evolved in three distinct phases. Initially hsp were investigated primarily as antigens, particularly when it was found that they were rather common targets of both the humoral and T cellmediated responses to intracellular pathogens like mycobacteria. Their recognition by T cells in models of autoimmune disease, particularly arthritis and diabetes, gave rise to much speculation that immune responses initially directed against hsp from pathogens might cross-react with self antigens including hsp themselves [1,2]. Since hsp are often up-regulated at sites of inflammation this would provide opportunities for persistent stimulation of cross-reactive hsp-specific T cells [3]. Such speculation continues, although more recent evidence points to antiinflammatory properties of T cells which recognize self hsp rather than their ability to induce autoimmune disease [4].

A second involvement of hsp in T cell-mediated immunity was demonstrated by the pioneering studies of Srivastava and colleagues, who showed that the chaperone function of many hsp (their ability to bind to and protect other polypeptides) allows them to deliver tumour antigens very effectively to antigen presenting cells [5,6]. This delivery appears to be receptor mediated, and hsp receptors are now being characterized, such as CD91 which binds several different hsp including two members of the hsp90 family and hsp70 [7]. In addition, when fusion proteins are created which contain antigen and part of the sequence of hsp70, the gain in immunogenicity is very marked, especially for the induction of responses by CD8+ T cells [8,9]. This function is analogous to that reported for antigens coupled to fragments of C3 which are potent inducers of antibody responses [10]. Thus both C3 and hsp might be regarded as physiological adjuvants, and serve as links between the innate and the acquired immune systems.

This brings us to the third immunological role of hsp, namely their ability to stimulate cells of the innate immune system, particularly antigen presenting cells, though interactions with other myeloid cells and endothelial cells have also been described. The initial observations were that hsp, usually tested as recombinant proteins, could elicit production of cytokines such as IL-1 or TNF α from monocyte/macrophage cells and cell lines [11–16].

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Another such paper by Ueki *et al.* [17] appears in this issue of CEI and documents cytokine production by macrophages in response to human hsp60; interestingly the investigators' initial interest seems to have been in hsp60 from *Actinobacillus actinomycetem-comitans*, a bacterium implicated in periodontal disease, but this did not induce cytokine production. Their initial idea that the bacterial hsp might mediate periodontal disease was modified therefore to suggest that human hsp60 released in response to infection would contribute to bone loss. In view of the accumulating literature on intrinsic stimulatory properties of hsp, it is timely to review current evidence on this novel feature of hsp.

A brief summary of the effects which have been measured and the hsp which have been implicated is shown in Table 1; this is not exhaustive and not all hsp have been tested in every system. The table also lists hsp by family but this conceals species differences reported by several workers; as noted already Ueki et al. [17] described a response mediated by human hsp60 but not hsp60 from two oral pathogens. Other workers have reported no effect of mycobacterial hsp60 in a bone resportion assay, but activity from E. coli hsp60 (GroEL). These results are surprising because in other studies mycobacterial hsp60 has been reported to be effective in producing effects on endothelial cells, and macrophages. This goes against the simple assumption that members of each hsp family interact with the same receptor on target cells, with variation in the effects observed reflecting the properties of the target cell. There are certainly difficulties in postulating receptors which can make relatively fine distinctions between members of the same hsp family from different species, given the conserved amino acid sequence and structure which is characterisic of hsp.

All workers in this field have had to contend with the suggestion that their hsp preparations, being derived from recombinant proteins, contain LPS (and other bacterial products), and that it is these which are responsible for cytokine induction or activation of myeloid and endothelial cells. Indeed, it is the case that most of the effects documented to occur with hsp can be duplicated by LPS. In addition the amounts of hsp required are usually in the $1-10\,\mu\text{g/ml}$ range whereas LPS produces similar effects at nanogram/ml concentrations. Thus there is considerable potential for artefact if careful controls are not included in experiments. The standard defence mounted by all investigators is to show that the effects of LPS in their assay system can be abolished by including polymyxin B (PmB) in cultures, whereas the effects of hsp are unaffected by PmB; conversely LPS is heat

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Table 1. Responses induced by heat shock protein families

Heat shock protein family	Target cell	Effect demonstrated
Hsp60	Monocyte/macrophages	Cytokine production: IL-1, TNFα, IL-6, IL-12 [24], IL-15, Nitric oxide production [13]
	Endothelial cells	Increased expression of CD62e, CD106, CD54 [14, 25]
	Vascular smooth muscle	IL-6 [14]
	Bone calvaria	Resorption – calcium release [26]
Hsp70	Monocyte/macrophages	Cytokine production: IL-1, TNFα [16]
Hsp90	Dendritic cells	Maturation: increased CD83, CD86
		Cytokine production: IL-12, TNF α , GM-CSF [27, 28]
Hsp27, Hsp10	Bone calvaria	Resorption – calcium release [26]

resistant whereas heating hsp preparations significantly decreases their activity.

However, it is still possible that these controls are not entirely adequate – what if hsp are up to their usual chaperoning tricks and are protecting small amounts of LPS and ensuring delivery to cellular receptors – much as they ensure and optimize delivery of antigenic peptides to antigen presenting cells. Under these circumstances the hsp would shield the LPS from binding to PmB, whilst destroying hsp by heating would remove their ability to transport small amounts of LPS. This is a crucial point which requires attention in the design and interpretation of future experiments. There is some evidence against the idea: Asea and colleagues showed that hsp70 induction of TNF α was blocked by a chelator of intracellular calcium whereas there was no such effect on the actions of LPS [16]. This clearly points to differing intracellular signalling mechanisms, but differences between signalling components utilized by hsp and LPS are conspicuously absent from the rest of the literature.

The same argument with respect to LPS could apply to other bacterial components which might find their way into recombinant preparations and interact with receptors of the innate immune system, particularly the Toll-like receptors (TLR). In addition to LPS which interacts with TLR4, bacterial peptidoglycans and lipoproteins interact with TLR2 [18], bacterial CpG DNA with TLR9 [19], flagellin with TLR5 [20], and viral doublestranded RNA with TLR3 [21]. This still leaves several additional TLR whose ligands are currently unknown but may well be products of bacteria.

There is now good evidence, from experiments in TLR4 deficient mice, that hsp60 requires TLR4 to elicit responses, and the same signalling molecules downstream of TLR4 (MyD88, TRAF6) which are critical in LPS signalling are also involved. It has also been suggested that TLR2 can also mediate responses to human hsp60 [22]; this would imply promiscuous association between hsp60 and two different TLRs - as was originally suggested for LPS before contaminants in LPS were identified which accounted for all of the response mediated through TLR2. Alternatively the observations would be explained if hsp60 were able to deliver LPS (or an alternative bacterial ligand) to TLR4, and a peptidoglycan-like entity to TLR2. Other aspects of the mechanism whereby LPS interacts with cells are worth recalling; the interaction is rather complex involving LPS binding protein which greatly increases the potency of LPS by enhancing delivery and binding to CD14. CD14 is indispensible for the LPS response, but seems to pass LPS on to TLR4 which is the signalling receptor. The TLR4 response in turn is significantly augmented by the molecule MD-2. This is produced by the responding cell and associates with TLR4, augmenting responses even when present at picomolar concentrations [23]. These two amplification mechanisms, LPS binding protein and MD-2, suggest that there might be scope for hsp to act in similar ways rather than as direct ligands of the signalling molecule, in addition to the postulated role of 'chaperoning' bacterial components to TLRs. It is unclear whether MD-2 is required for responses to hsp, and data on inhibition of responses to hsp by antibodies to CD14 are conflicting.

To clarify the direct stimulatory role of hsp several questions need to be answered. What is the nature of the interaction between hsp and TLRs, or other receptors such as CD91? Are these interactions specific and can they be mimicked by hsp-derived peptides? Ideally cocrystallization of hsp and receptor and direct measurement of affinity by surface plasmon resonance should be performed. TLR gene knock-out mice may also provide examples where responses to some hsp are maintained whilst those to bacterial components are lost. There is much work to do; delivery of antigenic peptides coupled with direct adjuvant function would represent a major physiological role for hsp in the initiation of immune responses, so we need to be confident of the mechanisms involved.

REFERENCES

- 1 van Eden W, Thole J, van der Zee R et al. Cloning of the mycobacterial epitope recognized by T lymphocytes in adjuvant arthritis. Nature 1988; 331:171–3.
- 2 Elias D, Marcus H, Reshef T, Ablamunits V, Cohen IR. Induction of diabetes in standard mice by immunization with the p277 peptide of a 60-kDa heat shock protein. Eur J Immunol 1995; 25:2851–7.
- 3 Lamb J, Young D. T cell recognition of stress proteins. A link between infectious and autoimmune disease. Mol Biol Med 1990; 7:311–21.
- 4 Anderton SM, van der Zee R, Prakken B, Noordzij A, van Eden W. Activation of T cells recognizing self 60-kD heat shock protein can protect against experimental arthritis. J Exp Med 1995; 181:943–52.
- 5 Blachere NE, Li ZH, Chandawarkar RY et al. Heat shock proteinpeptide complexes, reconstituted in vitro, elicit peptide-specific cytotoxic T lymphocyte response and tumor immunity. J Exp Med 1997; 186:1315–22.
- 6 Srivastava PK, Menoret A, Basu S, Binder RJ, McQuade KL. Heat shock proteins come of age: Primitive functions acquire new roles in an adaptive world. Immunity 1998; 8:657–65.
- 7 Basu S, Binder RJ, Ramalingam T, Srivastava PK. CD91 is a common receptor for heat shock proteins gp96, hsp90, hsp70, and calreticulin. Immunity 2001; 14:303–13.
- 8 Cho BK, Palliser D, Guillen E et al. A proposed mechanism for the

- induction of cytotoxic T lymphocyte production by heat shock fusion proteins. Immunity 2000; **12**:263–72.
- 9 Huang Q, Richmond JFL, Suzue K, Eisen HN, Young RA. In vivo cytotoxic T lymphocyte elicitation by mycobacterial heat shock protein 70 fusion proteins maps to a discrete domain and is CD4 (+) T cell independent. J Exp Med 2000; 191:403–8.
- 10 Dempsey PW, Allison MED, Akkaraju S, Goodnow CC, Fearon DT. C3d of complement as a molecular adjuvant: Bridging innate and acquired immunity. Science 1996; 271:348–50.
- 11 Friedland JS, Shattock R, Remick DG, Griffin GE. Mycobacterial 65-kD heat shock protein induces release of proinflammatory cytokines from human monocytic cells. Clin Exp Immunol 1993; 91:58-62.
- 12 Tabona P, Reddi K, Khan S et al. Homogeneous Escherichia coli chaperonin 60 induces IL-1 beta and IL-6 gene expression in human monocytes by a mechanism independent of protein conformation. J Immunol 1998; 161:1414–21.
- 13 Chen W, Syldath U, Bellmann K, Burkart V, Kolb H. Human 60-kDa heat-shock protein: a danger signal to the innate immune system. J Immunol 1999; 162:3212–9.
- 14 Kol A, Bourcier T, Lichtman AH, Libby P. Chlamydial and human heat shock protein 60s activate human vascular endothelium, smooth muscle cells, and macrophages. J Clin Invest 1999; 103:571–7.
- 15 Kol A, Lichtman AH, Finberg RW, Libby P, Kurt-Jones EA. Cutting edge: heat shock protein (HSP) 60 activates the innate immune response. CD14 is an essential receptor for HSP60 activation of mononuclear cells. J Immunol 2000; 164:13–7.
- 16 Asea A, Kraeft SK, KurtJones EA et al. HSP70 stimulates cytokine production through a CD14-dependant pathway, demonstrating its dual role as a chaperone and cytokine. Nature Med 2000; 6:435– 42
- 17 Ueki K, Tabeta K, Yoshie H, Yamazaki K. Self-heat shock protein 60 induces tumor necrosis factor-α in monocyte-derived macrophages: Possible role in chronic inflammatory periodontal disease. Clin Exp Immunol 2001, 126.

- 18 Takeuchi O, Hoshino K, Kawai T et al. Differential roles of TLR2 and TLR4 in recognition of gram- negative and gram-positive bacterial cell wall components. Immunity 1999; 11:443–51.
- 19 Wagner H. Toll meets bacterial CpG-DNA. Immunity 2001; 14: 499–502.
- 20 Hayashi F, Smith KD, Ozinsky A et al. The innate immune response to bacterial flagellin is mediated by Toll-like receptor 5. Nature 2001; 410:1099–103.
- 21 Alexopoulou L, CzopikHolt A, Medzhitov R, Flavell RA. Recognition of double-stranded RNA and activation of NF-κB byToll-like receptor 3. Nature 2001; 413:732–8.
- 22 Vabulas RM, AhmadNejad P, daCosta C et al. Endocytosed HSP60s use toll-like receptor 2 (TLR2) and TLR4 to activate the Toll/interleukin-1 receptor signaling pathway in innate immune cells. J Biol Chem 2001; 276:31332–9.
- 23 Visintin A, Mazzoni A, Spitzer JA, Segal DM. Secreted MD-2 is a large polymeric protein that efficiently confers lipopolysaccharide sensitivity to Toll-like receptor 4. Proc Natl Acad Sci USA 2001; 98:12156–61.
- 24 Breloer M, Dorner B, More SH, Roderian T, Fleischer B, vonBonin A. Heat shock proteins as 'danger signals': eukaryotic Hsp60 enhances and accelerates antigen-specific IFN-gamma production in T cells. Eur J Immunol 2001; 31:2051–9.
- 25 Verdegaal EME, Zegveld ST, van furth R. Heat shock protein 65 induces CD62e, CD106, and CD54 on cultured human endothelial cells and increases their adhesiveness for monocytes and granulocytes. J Immunol 1996; 157:369–76.
- 26 Nair S, Meghji S, Reddi K, Poole S, Miller A, Henderson B. Molecular chaperones stimulate bone resorption. Calcif Tissue Int 1999; 64:214–8.
- 27 SinghJasuja H, Scherer HU, Hilf N et al. The heat shock protein gp96 induces maturation of dendritic cells and down-regulation of its receptor. Eur J Immunol 2000; 30:2211–5.
- 28 Binder RJ, Anderson KM, Basu S, Srivastava PK. Cutting edge. Heat shock protein gp96 induces maturation and migration of CD11c (+) cells in vivo. J Immunol 2000; 165:6029–35.